UK Health Security Agency

Store LENTICULE disc samples @ -20°C± 5°C

Food and Environmental Proficiency Testing Unit

Laboratory Identification no. (Check) Lab No.

Dispatch date: **30 January 2023** Final date for return of results: **24 February 2023**

RESULTS TO BE RETURNED VIA ONLINE SURVEY AS PER EMAIL SENT TO YOU

Contact details:
The Organisers - FEPTU
UK Health Security Agency
61 Colindale Avenue,
London, NW9 5EQ, UK.
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UKAS PROFICIENCY TESTING
0006

Shiga toxin producing *Escherichia coli* (STEC) Request/Report Form	
Distribution No.: STX13	**Sample numbers**: **STX025 & STX026**
Download the sample Instruction sheet	www.gov.uk/government/publications/shiga-toxin-escherichia-coli-scheme-sample-instruction-sheet
Download the safety data sheet:	www.gov.uk/government/publications/safety-data-sheet-for-lenticules
If you cannot examine any of these samples return your results as 'Not examined'	
Request:	I. Examine samples for STEC genes

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SAMPLE: STX025

STEC / VTEC / EHEC (Shiga / verocytotoxic / enterohemorrhagic) testing:-

	Target Genes	Positive (circle)	CT value
STEC virulence gene detection	*stx* (*stx* 1/2 combined)	Yes / No / NE	
	stx 1	Yes / No / NE	
	stx 2	Yes / No / NE	
	eae	Yes / No / NE	
	Other ……..	Yes / No / NE	

	Not examined	Serogroup (e.g. O157)	Positive (circle)	CT value	Serotype (e.g. H7)	Positive (circle)	CT value
STEC identification gene detection	☐	O157	Yes / No / NE			Yes / No / NE	
		O26	Yes / No / NE			Yes / No / NE	
		O103	Yes / No / NE			Yes / No / NE	
		O104	Yes / No / NE			Yes / No / NE	
		O111	Yes / No / NE			Yes / No / NE	
		O145	Yes / No / NE			Yes / No / NE	
		Other…… ….	Yes / No / NE			Yes / No / NE	

Interpretation of results:-

Select interpretation of your results		Explanation
☐	Not examined / Not applicable	
☐	STEC not detected in the test portion of x g or xml	
☐	Presumptive detection of STEC in the test portion of x g or x ml	
☐	Presumptive detection of STEC causing the attaching and effacing lesion in the test portion of x g or x ml	
☐	Presumptive detection of STEC of xx serogroup in the test portion of x g or x ml	
☐	Other:	

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SAMPLE: STX026

STEC / VTEC / EHEC (Shiga / verocytotoxic / enterohemorrhagic) testing:-

	Target Genes	Positive (circle)	CT value
STEC virulence gene detection	*stx* (*stx* 1/2 combined)	Yes / No / NE	
	stx 1	Yes / No / NE	
	stx 2	Yes / No / NE	
	eae	Yes / No / NE	
	Other ……..	Yes / No / NE	

	Not examined	Serogroup (e.g. O157)	Positive (circle)	CT value	Serotype (e.g. H7)	Positive (circle)	CT value
STEC identification gene detection	☐	O157	Yes / No / NE			Yes / No / NE	
		O26	Yes / No / NE			Yes / No / NE	
		O103	Yes / No / NE			Yes / No / NE	
		O104	Yes / No / NE			Yes / No / NE	
		O111	Yes / No / NE			Yes / No / NE	
		O145	Yes / No / NE			Yes / No / NE	
		Other…… ….	Yes / No / NE			Yes / No / NE	

NE = Not Examined

Interpretation of results:-

Select interpretation of your results		Explanation
☐	Not examined / Not applicable	
☐	STEC not detected in the test portion of x g or xml	
☐	Presumptive detection of STEC in the test portion of x g or x ml	
☐	Presumptive detection of STEC causing the attaching and effacing lesion in the test portion of x g or x ml	
☐	Presumptive detection of STEC of xx serogroup in the test portion of x g or x ml	
☐	Other:	

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Any further comments:-

Authorised by:

Date reported:

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METHOD QUESTIONNAIRE – not all findings will be presented in the scheme report (include as much detail as possible or references):

1. Do you exactly follow ISO/TS 13136:2012 *Microbiology of food and animal feed -- Real-time polymerase chain reaction (PCR)-based method for the detection of food-borne pathogens - Horizontal method for the detection of Shiga toxin-producing* Escherichia coli *(STEC) and the determination of O157, O111, O26, O103 and O145 serogroups*	**YES**	**NO**

2. If the answer to question 1 is NO please state deviations to ISO/TS 13136:2012	
3. For routine samples, briefly describe your enrichment process e.g. 25 g into 225 ml mTSB+N/BPW 41.5 °C for 24hr	
4. Provide the details of your DNA extraction kit Include company name, kit name, protocol(s) if applicable: e.g., Promega, Maxwell® 16 Cell DNA purification kit	
5. If your DNA extraction process requires a platform please state platform used	

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6. State if you use conventional PCR or real-time PCR	

7. If using a <u>commercial kit</u> for primer / probe / amplification mix Please provide company name, kit name: e.g., Bio-Rad, IQ-Check STEC SerO; Bio-Rad IQ-Check® STEC VirX	

8. What volume of extracted DNA is used?	

9. Please complete the Cycle information tables

Pre-incubation:

Time (hh:mm:ss)		**Temperature (°C)**	

Cycle information:

Parameter	**Denaturisation**	**Cycling**				**Cooling**
Cycles	**x1**	**x**……..				**x1**
Temperature (°C)		**Step 1:**	**Step 2:**	**Step 3:**	**Step 4:**	
Hold (hh:mm:ss)						

Other:

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10. Amplification platform used e.g. Bio-Rad CFX96 Touch™ Deep Well RT-PCR Detection Systems	

11. Please use this space for any additional comments you feel are relevant or general comments about the scheme	